

YELLOW LUPIN TRANSCRIPTOME SEQUENCING TOWARDS IDENTIFICATION OF GENES ASSOCIATED WITH RESISTANCE TO *FUSARIUM SP.*

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RNA-SEQ

RNA-seq experiment was performed for **four yellow lupin genotypes (two resistant and two susceptible)**. Each genotype was grown in **two distinct environments: multiannual lupin monoculture (infected plants) and control field with crop rotation (non-infected plants)** (Table 1). The experiment was carried out in two biological replicates resulting in the total of **16 lines incorporated in the RNA-Seq analysis**.

Total RNA was extracted from leaves with aid of SV Total RNA Isolation System (Promega). TruSeq RNA Sample Preparation Kit (Illumina) was used to **cDNA libraries construction**. Sixteen RNA-seq libraries were sequenced using the **Illumina NGS platform (2x75 bp PE, HiSeq 1500)**. **A minimum of 59 million high quality sequence reads were obtained for each of biological replicates**.

TRANSCRIPTOME ASSEMBLY

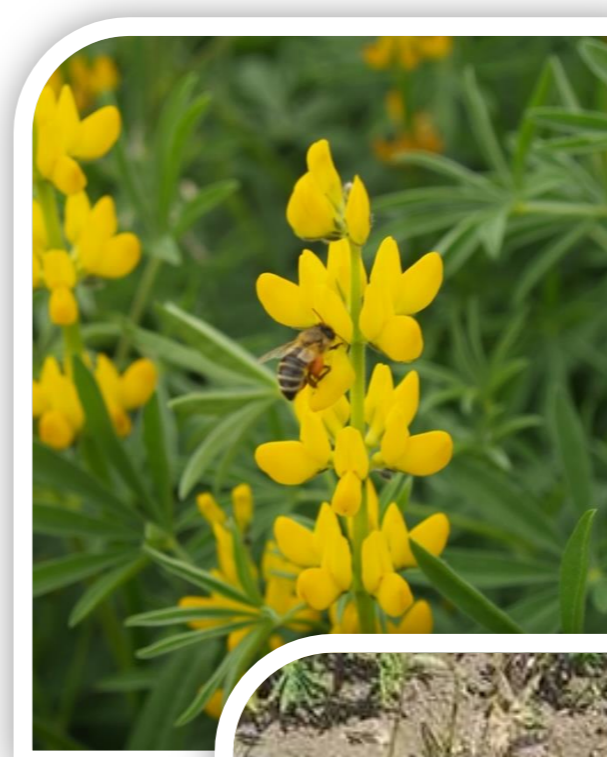
The reference transcriptome was **de novo assembled** on the basis of Lord and Perkoz libraries, using CLC Genomics Workbench 8.0.3 software. **We received 55043 contigs of an average length of 1058 bp (N50 = 1497 bp)**. Summary of contig measurements and contig length distribution are presented in Table 2 and Figure 1, respectively, while nucleotide distribution is shown on Figure 2.

Table 1. RNA-Seq experiment scheme.

Resistant genotypes	Susceptible genotypes
Lord/I	Perkoz /I
Lord/NI	Perkoz/NI
Z-505/3/I	Z-505/16/I
Z-505/3/NI	Z-505/16/NI

* I – infected plants, NI – non-infected plants

LUPINUS LUTEUS L.



Yellow lupin is one of three lupin crops. Lupins farming gained much attention recently, due to the fact that **lupin seeds are a valuable protein source**. On the other hand a serious threat in yellow lupin cropping are **fungal diseases e.g. fusariosis and anthracnose**, which may result in the total destruction of plantation. Up until now, only **one gene *Fus1* underlying resistance to *Fusarium sp.* has been identified** and used in the yellow lupin breeding programs, however its **molecular function has not been yet determined**.

Table 2. Contig measurements.

	Length
N50	1497
Minimum	298
Maximum	12264
Average	1058
Count	55043

This is the first study to investigate the molecular background of resistance to fusariosis in yellow lupin on the basis of RNA-Seq results. Transcriptome sequencing data serves as a basis in the reference transcriptome de novo assembly and differential gene expression analysis to identify candidate genes involved in the resistance to *Fusarium sp.*

Figure 1. Contig length distribution.

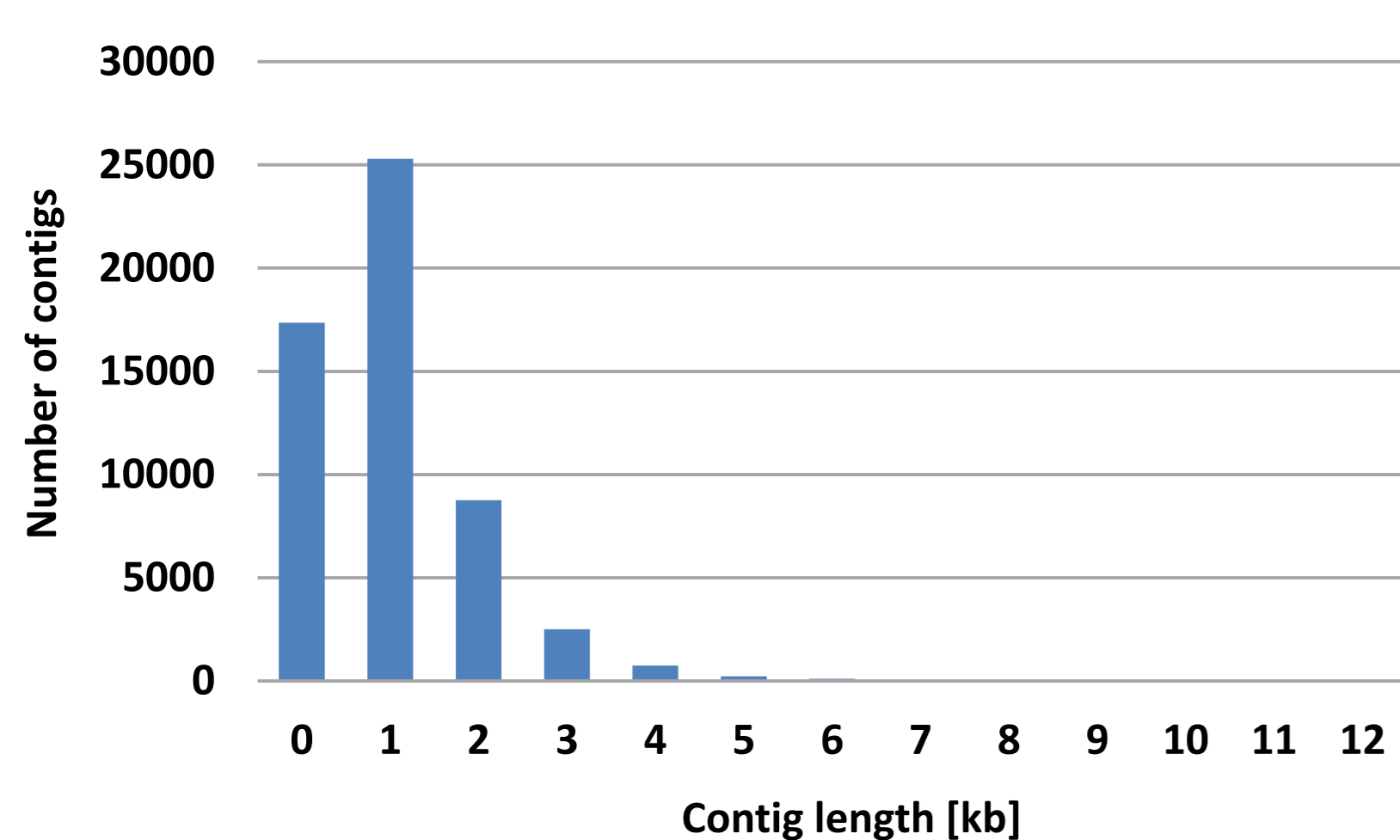


Figure 2. Nucleotide distribution in yellow lupin transcriptome assembly.

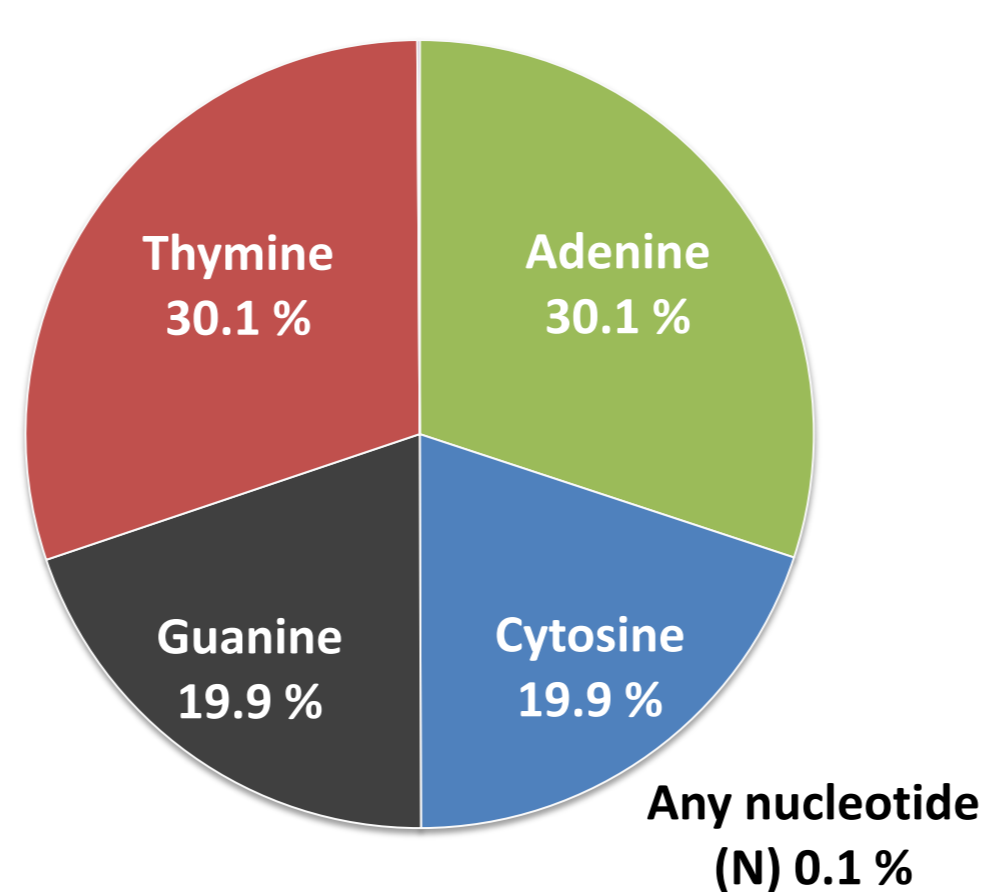
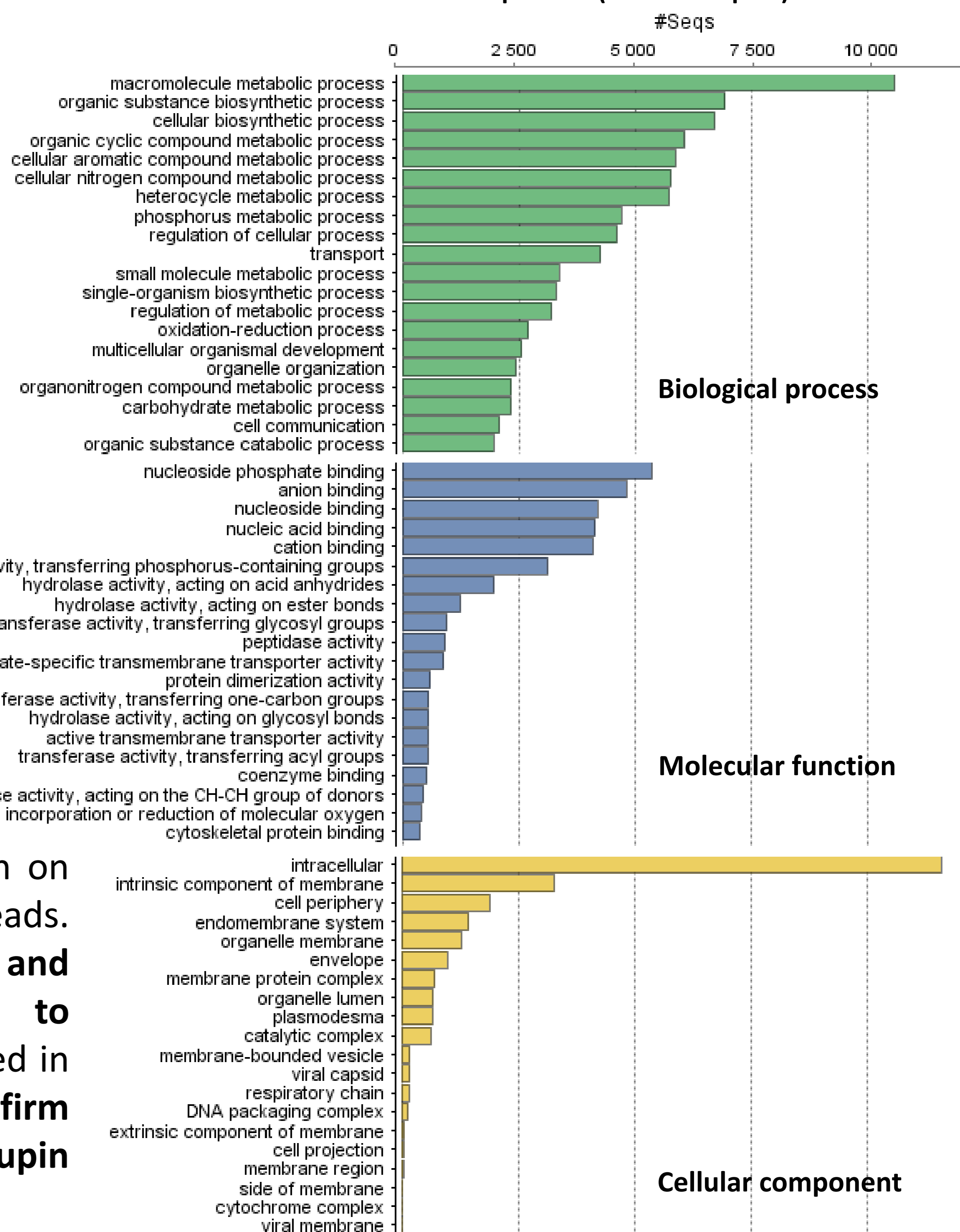


Figure 3. GO terms distribution according to biological processes, molecular function and cellular component (level 4 – top 20)



FUNCTIONAL ANNOTATION

Functional annotation of the transcriptome assembly was conducted using Blast2GO bioinformatic platform. **Terms of Gene Ontology (GO)** were distributed according to **biological processes, molecular function and cellular component** (Figure 3).

DIFFERENTIAL EXPRESSION

Gene expression levels were calculated using CLC Genomics Workbench on the basis of the assembled reference transcriptome and RNA-Seq short reads. **Comparison of type and abundance of transcripts from resistant and susceptible plants revealed differentially expressed genes belonging to a range of functional categories**. Candidate genes potentially involved in resistance to *Fusarium sp.* will be incorporated in **qPCR analyses to confirm differential expression in the wider group of susceptible and resistant lupin genotypes**.

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